File number: Dnr 2023/03884

Reference Material Datasheet

Version: 1.0

Issue date: 2023-10-09

Designation: RM Food 2023:P-CS

Batch no: 383

Date of production: 2023-02-01

Manufacturer: Swedish Food Agency, Sweden

-18 °C or lower (but not lower than -55 °C) Storage:

Batch expiry date: 2024-12-31

Manufacturer and contact information

Swedish Food Agency				
Company name	Swedish Food Agency (Livsmedelsverket)			
Department	Unit for Microbiology			
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Post code	SE-75126			
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Country	Sweden			
Telephone	+46 18 17 55 00			
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Intended use

This reference material is designed for internal quality control of analytical work at food microbiology laboratories. After reconstitution, the test material can be used for control of quantitative food microbiology analyses, as well as for direct or indirect quality control of microbiological media.

Content

Table 1. Microorganisms included in RM Food 2023:P-CS

Microorganism	Strain*	
Campylobacter jejuni	SLV-540	
Salmonella Enteritidis	SLV-436	
Staphylococcus saprophyticus	SLV-013	
Escherichia coli	SLV-165	

^{*} Internal strain identification number, Swedish Food Agency

Quality control

The reference material has been tested for homogeneity at the Swedish Food Agency. No statistically relevant difference has been observed between vials.



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Property values

Table 2. Quality control of RM Food 2023:P-CS. The results are from analysis of 10 individual vials, and are valid for a reconstitution volume of 104 ml. All values are expressed in log₁₀ cfu ml⁻¹.

Analysis	X _{RM}	S _{RM}	U RM	Acceptance limits	Method
Aerobic microorganisms	5.13	0.03	0.25	4.63 – 5.64	NMKL 86:2013
Thermotolerant Campylobacter	3.18	0.08	0.36	2.46 - 3.90	NMKL 119:2007
Salmonella	2.29	0.07	0.26	1.77 – 2.81	NMKL 71:1999

 x_{RM} : Property value, to be used for start-up control chart.

SRM: Standard deviation of the property value, can be used for start-up control chart.

 $u_{\rm RM}$: Standard uncertainty of the property value (includes uncertainty contributions from characterisation, homogeneity, transportation and method differences).

The lower/upper acceptance limits are calculated as: $x_{RM} \pm 2 * u_{RM}$ (expanded uncertainty at a 95 % confidence interval, with k = 2)

Traceability

Homogeneity, property values, standard deviations and control limits are calculated in accordance with ISO 17034 and ISO Guide 35. All values are metrologically traceable to the respective strains in the Swedish Food Agency's internal culture collection (Table 1).

Preparation of simulated food sample

Reconstitute the vial content according to the instructions on the last page.

Please note that the final 104 ml corresponds to the undiluted sample to be analysed.

The Swedish Food Agency uses peptone water (0.1 %) with NaCl (0.85 %) as diluent.

Analyses

The analyses should be performed in accordance with the methods used by the individual laboratory.

Control charts

Instructions for the construction of control charts are available at our website:

www.livsmedelsverket.se/RM-micro

Approved by

Jonas Ilbäck

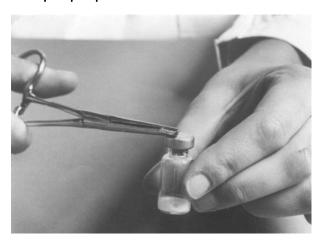
PT/RM Food Coordinator

Jonas Ilbäck



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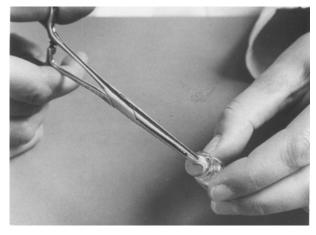
Sample preparation of freeze-dried cultures in glass vial (RM Food)



- **1**. Twist the flap on the aluminium cap in the direction of the arrow.
- 2. Remove the aluminium cap.



- **5**. Add 1 ml diluent with a sterile pipette.
- 6. Let the content dissolve (1-5 minutes).
- **7.** Using a sterile Pasteur pipette, transfer the suspension to a sterile bottle containing 100 ml room temperature diluent.
- **8.** Add another 1 ml and carefully rinse the walls of the vial with the Pasteur pipette.



- 3. Remove the rubber plug.
- **4.** Carefully burn the opening of the vial over a gas flame.



- **9.** Transfer the suspension to the bottle containing 100 ml diluent.
- **10.** Repeat steps 8 and 9 two more times with the same Pasteur pipette.
- **11.** After thorough intermittent mixing, the 104 ml sample is ready for analysis.
- 12. Perform the analyses immediately.*
- * For the majority of analyses, the final 104 ml sample is normally viable for up to 60 minutes. From experience however, lower results are often obtained—especially for Gram-negative microorganisms when the time between sample preparation and analysis is too long. As a general rule, it is therefore recommended to begin the analysis immediately after sample preparation.